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Genetic Variation among Endosymbionts of Widely Distributed Vestimentiferan Tubeworms

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Vestimentiferan tubeworms thriving in sulfidic deep-sea hydrothermal vents and cold seeps are constrained by their nutritional reliance on chemoautotrophic endosymbionts. In a recent phylogenetic study using 16S ribosomal DNA, we found that endosymbionts from vent and seep habitats form two distinct clades with little variation within each clade. In the present study, we used two different approaches to assess the genetic variation among biogeographically distinct vestimentiferan symbionts. DNA sequences were obtained for the noncoding, internal transcribed spacer (ITS) regions of the rRNA operons of symbionts associated with six different genera of vestimentiferan tubeworms. ITS sequences from endosymbionts of host genera collected from different habitats and widely distributed vent sites were surprisingly conserved. Because the ITS region was not sufficient for distinguishing endosymbionts from different habitats or locations, we used a DNA fingerprinting technique, repetitive-extragenic-palindrome PCR (REP-PCR), to reveal differences in the distribution of repetitive sequences in the genomes of the bacterial endosymbionts. Most of the endosymbionts displayed unique REP-PCR patterns. A cladogram generated from these fingerprints reflected relationships that may be influenced by a variety of factors, including host genera, geographic location, and bottom type.

Invertebrates that are endemic to the highly sulfidic, reducing environments at deep-sea hydrothermal vents and cold seeps are commonly associated with chemosynthetic endosymbiotic bacteria (7, 18, 19). These bacteria oxidize the reduced sulfur compounds that are abundant in hydrothermal fluid (8, 36, 46). The resultant energy produced by the endosymbiont is coupled to the production of carbon sources that support the growth and maintenance of the invertebrate hosts (6, 8, 37, 48). The metabolic link between the invertebrate hosts and their endosymbionts has obvious implications for the dispersal and colonization strategies employed by the host organisms.

Vest endemic host organisms may employ one of three different mechanisms for transmission of their endosymbionts to the next generation. Previous studies have shown that the Vesicomyid bivalves form species-specific associations with endosymbiotic bacteria (12, 13, 16). Host specificity is maintained in these associations by vertical transmission of the bacteria through the egg from the parent to the offspring (3, 35). The endosymbiont and host phylogenies are congruent, which is consistent with a vertical mode of symbiont transmission (3, 4, 12, 35). In contrast, results from several studies suggest that endosymbiont transmission in vestimentiferan tubeworms occurs horizontally (5, 17), possibly through ingestion of bacteria upon larval settlement (28, 42). It is also possible that horizontal transmission in vestimentiferan-bacterial symbioses could occur through infection of new recruits with bacteria previously associated with established congeners (20), although potential mechanisms for this transfer have not been determined.

If horizontal transmission is indeed the mechanism for bacterial acquisition in vestimentiferans, we expect that biogeographic variation would exist among symbionts acquired from the same host species collected in geographically isolated vent and seep sites. Conversely, if vestimentiferan larvae settle in the same locations where their symbionts are acquired, we expect that vestimentiferans living at the same site would harbor identical symbionts.

In a recent study, genetic variation in vestimentiferan endosymbionts was investigated by comparing sequences of the 16S rRNA genes of bacteria collected from various species of hydrothermal vent and cold seep tubeworms (20). The results showed a marked phylogenetic distinction between vestimentiferan endosymbionts collected from hydrothermal vents and soft-bottom, cold seep environments. This study suggested that vestimentiferans acquire one of two distinct species of free-living bacteria depending on whether they settle on basaltic, hydrothermal vent sites or sedimentioned, cold seep sites. A third, unique bacterial species was discovered in a tubeworm that inhabited a sedimented whale fall. Significant genetic variation was not found among symbionts found within each of the habitat types. Symbionts from three different host genera collected from five different hydrothermal vent sites along the East Pacific Rise (EPR) had identical 16S rRNA gene sequences.

The observation of so little sequence variation among endosymbiont 16S rRNA genes suggests that all tubeworm endosymbionts may belong to a single species. However, the conservative nature of the 16S rRNA gene often renders it inadequate for distinguishing among conspecific strains of bacteria (50). Therefore, it is not surprising that Feldman et al. (20) did not find significant genetic variation within the habitat clades. It is possible that significant genetic variation exists...
between endosymbionts of geographically distributed vestimentiferan hosts but that these species- or strain-level differences have not been detected by approaches that have been employed to date.

The goal of our study was to resolve the question of genetic variation among widely distributed vestimentiferan endosymbionts by utilizing two molecular techniques that have historically proven useful for comparing closely related strains of bacteria. Our first approach entailed sequencing of the internal transcribed spacer (ITS) region, a noncoding region within the rRNA that often contains genetic variation sufficient for differentiating species of prokaryotes (2). Our second approach involved using a DNA fingerprinting technique that utilizes repetitive extragenic palindromes (REPs) as priming sites to generate diagnostic banding patterns from prokaryotic genomic DNA (25, 45). The REP-PCR technique is particularly useful for investigations of bacterial symbiosis because REP sequences have been found only in bacterial DNA; phage and eukaryotes do not contain REP elements (25). By virtue of these repetitive elements, symbiont DNA can be selectively amplified from a mixed population of symbiont and host DNA to allow analysis of bacterial population diversity. Our results provide information on the extent of genetic variation among the symbionts, as well as further evidence to support a horizontal model of symbiont acquisition among the vestimentiferan tubeworms.

**MATERIALS AND METHODS**

**Sample collection.** Vestimentiferans were collected with the aid of submarines from several locations, including two sites in the Western Pacific Ocean, four sites along the EPR, two sites in the Pacific Northwest, one site along the Galapagos Rift (GR), two sites along the western coast of California and Mexico, and two locations in the Gulf of Mexico (GOM) (Table 1). Samples included in this study were chosen to represent the entire range of host vestimentiferan distribution. Habitat types included hydrothermal vent sites with a vesitimentiferan host was aseptically removed, and a small portion of it was homogenized in 5 M guanidinium isothiocyanate. Prior to DNA extraction, 25 μl of 5 M guanidinium isothiocyanate homogenate was added to the sample and incubated at 65°C for 1 h at 4°C. The samples were briefly spun to remove the Chelex 100. Total bacterial DNA was extracted from 100 μl of guanidinium isothiocyanate homogenate using the IsoQuick DNA purification kit (Orca, Bothell, Wash.) according to the manufacturer’s instructions and quantified spectrophotometrically. In addition to symbiont DNA, host DNA from three different genera was extracted from ventenousm smear (non-symbiont-containing tissue) using the same extraction protocol to serve as controls. A number of the DNA samples used in this study (e.g., 18°S Riftia pachyptila, Guaymas Vent and Seep [GY-V and GY-S, respectively] Escarpita spicata, whale E. spicata, Gorda Lamellibrachia, sp. 21°N Oasia alvinae; and Gy-V R. pachyptila) were prepared separately at Rutgers University according to previously published protocols (20).

**16S rRNA gene characterization.** The methods used to PCR amplify, purify, and sequence the 16S rRNA symbiont genes were as described in the work of Feldman et al. (20). DNA sequence alignments were initially constructed using PileUp and then optimized (minimizing overall alignment differences) by eye in the SeqLab environment (Genetics Computer Group). Insertions and deletions were eliminated from the alignment before phylogenetic trees were determined. Phylogenetic trees for the symbiont 16S rRNA genes were computed using fastDNAml and bootstrapped 100 times using fastDNAml boot (21, 34). For the maximum likelihood analyses, operational taxonomic unit input order was randomized and global rearrangements were performed at each bootstrap replication.

**ITS region characterization.** PCRs amplified the ITS region of the symbiont ribosomal DNA (rDNA) using two ITS-specific primers, ITS16F-G1 (5′-AGAG TCCTAACAC-G-3′) (27) and ITS23R-L1 (5′-CAACGACATCCACCGT-3′) (27). ITS16F-G1 is nested about 30 to 40 nucleotides upstream from the spacer boundary in the 3′ end of the 16S rRNA gene (positions 1491 to 1505, Escherichia coli 16S rRNA gene). ITS23R-L1 is nested in the 5′ end of the 23S rRNA gene, approximately 20 bases downstream from the spacer boundary (positions 21 to 35, E. coli 23S rRNA gene). PCR mixtures (50-μl total volume) contained final concentrations of the following: 50 ng of symbiont DNA, 1× PCR buffer, 0.2 mM deoxynucleoside triphosphates (dNTPs), 1.5 mM MgCl2, 5% (vol/vol) acetamide, and 1.25 U of Taq polymerase (Promega). PCR conditions were as follows: 35 cycles, each consisting of denaturation at 92°C for 1 min, hybridization at 55°C for 2 min, and elongation at 72°C for 2 min with 5 s added to each extension per cycle. A hot start (9) with denaturation at 95°C for 2 min was used at the beginning of the reaction to heighten reaction specificity. In addition, acetamide was added to the PCR to encourage more efficient amplification of GC-rich templates (38). All PCRs were performed on an MJ Minicycler (MJ Research, Inc., Watertown, Mass.).

Because our attempts to directly sequence the smaller ITS PCR amplicon yielded poor results, amplified ITS regions were cloned to facilitate sequencing. Amplification products were pooled from three separate PCRs and cloned directly using the TA cloning kit with the PCR-II or the PCR-2.1-TOPO cloning vectors (Invitrogen, San Diego, Calif.) according to the manufacturer’s instructions. Plasmid preparations were made using a standard alkaline-lysis preparation (39). The sizes of the inserts were verified before sequencing by restriction analysis. The plasmid DNA was further purified for sequencing using the Plasmid Miniprep (Qiagen, Inc., Valencia, Calif.) according to the manufacturer’s instructions. Two clones from each amplified ITS region were sequenced using the Perkin Elmer (Foster City, Calif.) ABI BigDye dye-termination cycle sequencing ready reaction kit with Ampli Taq DNA polymerase FS according to the manufacturer’s instructions. Sequencing was performed on an ABI PRISM 310 genetic analyzer.

The ITS regions were bidirectionally sequenced and confirmed prior to their alignment using the Sequence Navigator and AutoAssembler programs (Applied BioSystems, Inc.). Only sequences that overlapped with ≥95% identity were included in the analysis. Final alignments were generated in the Genetic Data Environment, version 3.2. Phylogenetic relationships were determined from

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**TABLE 1. Collection sites of vestimentiferan symbionts included in this study**

<table>
<thead>
<tr>
<th>Host species</th>
<th>Collection site</th>
<th>Latitude; longitude</th>
<th>Substrate, community type</th>
<th>Depth (m)</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>R. pachyptila</em></td>
<td>18°S, EPR (18S)</td>
<td>18°36′S; 113°24′W</td>
<td>Basaltic, vent</td>
<td>2,637</td>
<td>20</td>
</tr>
<tr>
<td><em>R. pachyptila</em></td>
<td>9°N, EPR (9N)</td>
<td>9°49′N; 104°17′W</td>
<td>Basaltic, vent</td>
<td>2,516</td>
<td>20</td>
</tr>
<tr>
<td><em>R. pachyptila</em></td>
<td>Guaymas Basin vent (GY-V)</td>
<td>26°59′N; 111°24′W</td>
<td>Basaltic, vent</td>
<td>2,016</td>
<td>20</td>
</tr>
<tr>
<td><em>R. pachyptila</em></td>
<td>GR</td>
<td>26°48′N; 80°13′N</td>
<td>Basaltic, vent</td>
<td>2,451</td>
<td>20</td>
</tr>
<tr>
<td><em>R. pachyptila</em></td>
<td>13°N, EPR (13N)</td>
<td>12°49′N; 103°56′W</td>
<td>Basaltic, vent</td>
<td>2,630</td>
<td>This study</td>
</tr>
<tr>
<td><em>Oasia alvinae</em></td>
<td>21°N, EPR (21N)</td>
<td>20°47′N; 109°08′W</td>
<td>Basaltic, vent</td>
<td>2,577</td>
<td>20</td>
</tr>
<tr>
<td><em>T. jerichonana</em></td>
<td>13°N, EPR (13N)</td>
<td>12°49′N; 103°56′W</td>
<td>Basaltic, vent</td>
<td>2,630</td>
<td>This study</td>
</tr>
<tr>
<td><em>T. jerichonana</em></td>
<td>9°N, EPR (9N)</td>
<td>9°49′N; 104°17′W</td>
<td>Basaltic, vent</td>
<td>2,516</td>
<td>20</td>
</tr>
<tr>
<td><em>Ridgea piscesae</em></td>
<td>JDF</td>
<td>45°58′N; 130°01′W</td>
<td>Basaltic, vent</td>
<td>1,550</td>
<td>This study</td>
</tr>
<tr>
<td>Lamellibrachia sp.</td>
<td>Green Canyon, GOM</td>
<td>27°41′N; 19°32′W</td>
<td>Sedimented, seep</td>
<td>700</td>
<td>5</td>
</tr>
<tr>
<td>Lamellibrachia sp.</td>
<td>Gorda Ridge (Gorda)</td>
<td>42°45′N; 120°42′W</td>
<td>Sedimented, seep</td>
<td>2,847</td>
<td>20</td>
</tr>
<tr>
<td>E. laminata</td>
<td>WFE</td>
<td>26°02′N; 84°55′W</td>
<td>Sedimented, seep</td>
<td>3,243</td>
<td>20</td>
</tr>
<tr>
<td>E. spicata</td>
<td>Santa Catalina Basin (whale)</td>
<td>33°12′N; 118°30′W</td>
<td>Sedimented, whale fall</td>
<td>1,240</td>
<td>20</td>
</tr>
<tr>
<td>E. spicata</td>
<td>Guaymas Basin vent (GY-V)</td>
<td>26°59′N; 111°24′W</td>
<td>Basaltic, vent</td>
<td>2,016</td>
<td>This study</td>
</tr>
<tr>
<td>E. spicata</td>
<td>Guaymas Basin seep (GY-S)</td>
<td>26°59′N; 111°24′W</td>
<td>Sedimented, seep</td>
<td>2,016</td>
<td>This study</td>
</tr>
<tr>
<td>L. columna</td>
<td>Lau Basin, Hine Hina (Lau-Fiji)</td>
<td>22°32′S; 170°43′W</td>
<td>Sedimented, seep</td>
<td>1,859</td>
<td>42</td>
</tr>
<tr>
<td>Undescribed species</td>
<td>Nikko Scamound (Nikko)</td>
<td>23°05′N; 142°20′E</td>
<td>Basaltic, vent?</td>
<td>433</td>
<td>This study</td>
</tr>
</tbody>
</table>

* See the work of Feldman et al. (20) for a map of the sample distribution.
bootstrap values of the method with 100 bootstrap replicates. Each number on a branch indicates the number of times (out of 100) that the node was supported by the bootstrap analysis. Only Lamellibrachia endosymbiont; M99446, *Thiobacillus hydrothermalis* and final concentrations of 1\(3^\text{mM} \times 10^{-3}\) (44). Each 25-\(\mu\)l REP-PCR mixture contained 50 ng of symbiont or host DNA and 0.5 unit of Taq polymerase (Gibco). One cycle of 95°C for 7 min was performed to denature the genomic DNA, followed by 30 cycles of 94°C for 1 min, 44°C for 1 min, and 65°C for 8 min, and a final extension at 65°C for 15 min. PCRs were added: GY-V, E. coli; U77478, 9N and 18S R. pachyptila endosymbiont and 9N T. jerichonana endosymbiond; U77479, GOM Lamellibrachia sp. endosymbiont and WFE E. lamintana endosymbiont; U77480, JDF Ridgea piceasce endosymbiont and GY-V and R. pachyptila endosymbiont; U77481, Lau-Fiji. L. columnna endosymbiont; U77482, whale E. spicata endosymbiont. (B) Dendrogram showing the relationships of vestimentiferan symbionts based on ITS sequences. The tree was rooted with the symbiont from a marine bivalve, L. floridana, as an outgroup. The tree was constructed using the neighbor-joining method with 100 bootstrap replicates. Each number on a branch indicates the number of times (out of 100) that the node was supported by the bootstrap analysis. Only bootstrap values of \(\pm 50\%\) are reported. Two major clusters of endosymbionts are indicated. Scale bar represents 10% sequence dissimilarity.

**RESULTS AND DISCUSSION**

16S rDNA sequence variation among vestimentiferan endosymbionts. In order to maintain a comparable data set, many of the endosymbiont DNA samples used in our ITS and REP-PCR analyses are the same DNA samples used in the previous 16S rDNA study of geographically distinct endosymbionts (20). Our study also includes additional samples to expand the representation of symbiont distribution. In concordance with our expanded data set, a new 16S rDNA phylogenetic tree was generated to include three of the endosymbionts that were added: GY-V E. spicata, GY-S E. spicata, and the undescribed species from the Nikko Seamount; and AF165908, E. spicata endosymbiont, GY-S.

**FIG. 1.** (A) Dendrogram based on 16S rDNA sequences of vestimentiferan symbionts. Phylogenetic relationships were computed using fastDNAml and bootstrapped 100 times using fastDNAml boot (21, 34). For the maximum likelihood analyses, operational taxonomic unit input order was randomized and global rearrangements were performed at each bootstrap replication. Two major groups of endosymbionts are indicated. Scale bar represents 10% sequence dissimilarity. Reference sequences used in this 16S study were obtained from the GenBank database and include the following accession numbers: L25712, Codakia costata symbiont; L25711, Anodontia phillipiana symbiont; M94946, *Cabytgona magnifica* symbiont; L25710, *Cabytgona pacifica* symbiont; L25718 and L25719, Cabytgona elongata symbiont; M90662, *Thiobacillus hydrothermalis*; V00348, E. coli; U77478, 9N and 18S R. pachyptila endosymbiont and 9N T. jerichonana endosymbiond; U77479, GOM Lamellibrachia sp. endosymbiont and WFE E. lamintana endosymbiont; U77480, JDF Ridgea piceasce endosymbiont and GY-V and R. pachyptila endosymbiont; U77481, Lau-Fiji. L. columnna endosymbiont; U77482, whale E. spicata endosymbiont. (B) Dendrogram showing the relationships of vestimentiferan symbionts based on ITS sequences. The tree was rooted with the symbiont from a marine bivalve, L. floridana, as an outgroup. The tree was constructed using the neighbor-joining method with 100 bootstrap replicates. Each number on a branch indicates the number of times (out of 100) that the node was supported by the bootstrap analysis. Only bootstrap values of \(\pm 50\%\) are reported. Two major clusters of endosymbionts are indicated. Scale bar represents 10% sequence dissimilarity.
symbionts is maintained between symbionts from the same host, *E. spicata*, collected from vent (GY-V) and seep (GY-S) habitats at a single site in Guaymas Basin. Likewise, symbionts from two different vestimentiferan hosts are closely related when collected from the same habitat type (e.g., 9N *R. pachyptila* and *T. jerichonana* symbionts, both from vent habitats).

**ITS sequence variation among vestimentiferan symbionts.**

ITS amplifications of vestimentiferan symbionts were generally 550 bp in length, although some individuals yielded a second, larger PCR product of roughly 800 bp in length (data not shown). The larger amplicon did not match any ITS sequences in the database, and we found that its intensity in the total PCR product was reduced by the use of higher annealing temperatures. For these reasons, we believe that the larger product represented an artifact of the PCR rather than rRNA operon heterogeneity. To separate these two distinct amplicons, we cloned the ITS PCR amplification products and used only the smaller amplicon from each symbiont for sequence analysis. As in many bacteria, all of the endosymbiont ITS regions contained putative sequences for the tRNA genes for alanine and isoleucine (Fig. 2).

Five of the symbionts contained prominent insertion sequences (approximately base position 312 in ITS region) that totaled approximately 50 bp (Fig. 2). The sequences of these insertions were nearly identical in all of the symbionts in which they were found.

Groups I and II in the 16S rDNA cladogram (Fig. 1A) correlate well with groups I and II in the ITS cladogram (Fig. 1B), with one exception: *E. laminata* symbiont, WFE, appears in group I according to its 16S rDNA sequence and in group II according to its ITS sequence (Fig. 1).

To determine if there was a significant correlation between ITS sequence diversity and either geography or habitat type, we performed two- and three-way Mantel tests to compare distance matrices generated from pairwise differences between ITS sequences, geographic distances, or habitat types.
results of these Mantel tests are shown in Table 2. Despite the remarkable similarity among the ITS sequences, the genetic distances among ITS regions correlate significantly with geographic distances. Although none of the correlations were particularly strong ($r \leq -0.334$ for all tests), $P$ values from two different Mantel tests indicated that ITS sequence similarity and geography were significantly correlated ($P = 0.0349$, test 3; $P = 0.0480$, test 4). In contrast, $P$ values for a two-way Mantel test of ITS sequence similarity versus habitat type alone (test 1) or a three-way test versus habitat type with geography factored out (test 5) were not statistically significant ($P = 0.146$ and 0.102, respectively). Thus, ITS sequence similarity was significantly correlated with broad-scale endosymbiont geography ($P \leq 0.05$) but not significantly correlated with habitat type. In light of the fact that the ITS sequences for 9°N, 18°S, and Guaymas R. pachyptila were identical, our results suggest that the ITS region is variable enough to show broad-scale biogeographic variation but is not variable enough to distinguish among symbionts located along the same ridge axis. These results are in contrast to the results of Feldman et al. (20) and the new analysis of 16S rDNA relationships presented in this study, which suggest that symbiont sequence distances correlate with habitat type (basaltic, vent versus sedimented, seep substrates). Although our data suggest that the geographic distribution of the hosts may be a dominant influence in determining genetic variation of the endosymbionts, it is clear that several factors affect endosymbiont diversity.

Historically, the ITS region has been useful for discriminating among strains of several bacterial species, including a hyperthermophilic archaean (11), Rhizobium sp. (30), Bifidobacterium sp. (31), infectious Pseudomonas sp. (43), and Trichodesmium sp. (49). In the present study, we found that the ITS region was much more conserved than expected. Although the ITS region is usually very informative at the subspecies and/or strain level (11, 30, 31, 49), it has been unreliable in some cases. In studies of the Mycobacterium tuberculosis complex (23) and with several Listeria isolates (15), the ITS region failed to differentiate between species and/or strains.

**REP-PCR fingerprints of the vestimentiferan symbionts.** The REP-PCR technique is particularly well suited for investigations of bacterial symbiosis, in that it allows investigators to survey the symbiont genome in the presence of host DNA because the REP priming sites are exclusively bacterial (25). To confirm that REP sequences were not present in the genomes of the vestimentiferan hosts, we performed REP-PCR amplifications on DNA that was aseptically acquired from vestimentum (non-symbiont-containing) tissue from three different host genera (*Escarpia*, *Riftia*, and *Tevnia*). Most of these samples failed to amplify at all (data not shown). The exception was *E. laminata*, which produced a single band that did not correspond with any of the bands seen in the REP fingerprint of its endosymbiont.

![Digital image of the REP-PCR fingerprint patterns of the vestimentiferan symbionts. Shown are the REP-PCR products generated by using chromosomal DNA of endosymbionts extracted from the following vestimentiferans: R. pachyptila, GY-V (lane 1); R. pachyptila, 9N (lane 2); R. pachyptila, GR (lane 3); R. pachyptila, 18S (lane 4); Ridgdea piscesae, JDF (lane 5); T. jerichonana, 9N (lane 6); undescribed sp., Nikko Seamount (lane 7); L. columnna, Lau-Fiji (lane 8); E. spicata, whale (lane 9); E. spicata, GY-V (lane 10); E. spicata, GY-S (lane 11); Lamelibrachia sp., GOM-10 (lane 12); E. laminata, WFE (lane 13); Lamelibrachia sp., GOM-11 (lane 14). Lanes H, HI Low DNA marker; lane M, 1-kb DNA ladder. Molecular sizes (base pairs) of selected bands in the DNA markers are indicated. The photo represents a composite of a larger gel that was altered in Adobe Photoshop version 5.0 to include only the individuals presented in this study.**

**TABLE 2.** Mantel tests of correlation among geographic, habitat, and ITS sequence similarity matrices for vestimentiferan symbionts

<table>
<thead>
<tr>
<th>Test</th>
<th>Matrix A</th>
<th>Matrix B</th>
<th>Conditional matrix C</th>
<th>$r$</th>
<th>$t$</th>
<th>$P$</th>
</tr>
</thead>
<tbody>
<tr>
<td>Two-way</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>ITS sim</td>
<td>Habitat</td>
<td>-0.133 - 1.402 0.1459</td>
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<td></td>
<td></td>
</tr>
<tr>
<td>2</td>
<td>Geography</td>
<td>Habitat</td>
<td>0.006 0.053 0.4555</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>ITS sim</td>
<td>Geography</td>
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<tr>
<td>Three-way</td>
<td></td>
<td></td>
<td></td>
<td></td>
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<td></td>
</tr>
<tr>
<td>4</td>
<td>Geography</td>
<td>ITS sim</td>
<td>-0.334 -2.349 0.0480</td>
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<td></td>
<td></td>
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<tr>
<td>5</td>
<td>Habitat</td>
<td>ITS sim</td>
<td>-0.139 -1.508 0.1019</td>
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</tr>
</tbody>
</table>

* $\sim$ similarity.
tom substrate types, clade together and group away from the symbionts collected from the Guaymas Basin vent (GY-V). Within the soft-bottom grouping of *Escarpia* symbionts, it appears that the symbionts from California and Guaymas (both on the western coast of North America) group away from the symbiont from Florida (on the eastern coast). The symbionts from the Nikko Seamount and Lau-Fiji Basin are clearly different from the rest of the symbionts, presumably because they are so geographically removed from the rest of the collection sites.

REP-PCR has been useful for genomic fingerprinting of various strains of bacteria, including *Actinobacillus* (1), *Rhizobium* (10, 30), *Legionella* (24), *Streptococcus* (44), *Bacillus* (26), and *Citrobacter diversus* (51) strains. In the present study, we found that the REP-PCR fingerprint analysis was sufficiently fine-scaled to reveal some interesting strain-level genetic variation among vestimentiferan symbionts. Presumably, this is because the REP analysis targets the whole genome, rather than a specific, relatively conservative region of the DNA. This physically maximizes the probability of finding strain-level variation. It should be noted that the symbiont of *O. alvinae* was not included in the REP analysis, because a reliable REP-PCR amplification and subsequent fingerprint pattern were not obtainable with this DNA. When the DNA was run on an agarose gel, it appeared sheared, and although it was sufficiently large enough for performance of the 550-bp ITS amplification, there was insufficient high-molecular-weight genomic DNA to support the REP analysis.

**Implications for symbiont relatedness and acquisition.** Previous studies have suggested that vestimentiferans acquire their symbionts through ingestion of free-living bacteria upon larval settlement (28, 42). If this mechanism alone is employed, one would expect that symbionts from two different vestimentiferan host species living at the same location would be identical or related more closely to each other than they are to other endosymbionts collected from distant vent sites. This, in fact, was not the case, as symbionts from 9°N *T. jerichonana* and 9°N *R. pachyptila* (both collected from the same rock) did not display identical ITS sequences or REP-PCR fingerprints. Instead, the symbionts grouped according to their respective host species as revealed through comparisons of shared bands in their REP-PCR fingerprints.

One explanation for this is that larvae could acquire their symbionts prior to attachment, either from a different substrate than where they eventually colonize or from the water column. This could enable two vestimentiferans that reside in the same site to harbor different symbionts. Alternatively, symbiont acquisition may be related to ecological succession. *T. jerichonana* is the first colonizer of new hydrothermal vent fields, in areas of the most intense diffuse flow (40). Presumably, this is because *T. jerichonana* is more tolerant (than *R. pachyptila*) of the high temperatures and elevated concentrations of H₂S that are present in newly formed vent sites. It is only after *T. jerichonana* has become established and the levels of reduced chemicals have attenuated that *R. pachyptila* begins to colonize a new hydrothermal vent site (40). The *T. jerichonana* and *R. pachyptila* hosts may simply acquire a different subset of a free-living chemosynthetic community due to temporal and/ or spatial consequences of ecological succession.

Specific host recognition mechanisms may exist that allow certain strains of endosymbiotic bacteria to survive in specific vestimentiferan hosts. Although these mechanisms have not been demonstrated in vestimentiferans, they could explain how two different hosts living on the same rock could harbor different symbionts. Alternatively, the larvae may acquire bacteria that have been released from congeners residing at the site of larval settlement, potentially through host decomposition. These mechanisms might yield REP-PCR fingerprint patterns that imply close relationships between endosymbionts of the same host genus. Because only one individual from each vestimentiferan genus was sampled from most of the locations, we cannot determine the extent of ITS sequence variation among endosymbionts at each site.

The extent of vestimentiferan symbiont relatedness and the host transmission mechanisms used to maintain these symbioses have remained ongoing questions. The results presented here demonstrate the existence of significant strain-level variation between endosymbionts of different vestimentiferan hosts collected from geographically separated areas. Further work to pinpoint the stage during vestimentiferan development when the symbiosis is first established will shed light on symbiont transmission and help to explain the genetic diversity among symbiont populations.

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